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# Synthesis of (±)-coerulescine and a formal synthesis of (±)-horsfiline

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Abstract—A straightforward synthesis of (±)-coerulescine and (±)-horsfiline has been established from 3-formyl-3-phenylpyrrolidine employing 4-hydroxypiperidine as the starting material. There are two remarkable steps for the synthesis of  $(\pm)$ -coerulescine and (±)-horsfiline. One is the rapid access to produce 3-formyl-3-phenylpyrrolidine by Lewis acid-catalyzed rearrangement of 3,4dihydroxy-4-phenylpiperidine. The other key step is an intramolecular electrophilic cyclization from 3-benzylcarbamoyl-3-phenylpyrrolidine to the 3,3-spirocyclic 2-oxindole ring skeleton. © 2005 Elsevier Ltd. All rights reserved.

#### 1. Introduction

The derivatives of spiropyrrolidinyl-oxindole skeleton alkaloid are represented in nature such as spirotryprostatins A and B, elacomine, and strychnophylline. Typically, they are biosynthesized from tryptamine or tryptophan derivatives and are characterized by a spiro-fused ring at the C-3 position.<sup>4</sup> The majority of these natural products have interesting biological activities and pharmacological properties. Among them, coerulescine (1) and horsfiline (2) represent the simplest prototype members of this subfamily. Recently, coerulescine (1) was isolated from the blue canary grass Phalaris coerulescens. 5a,b Horsfiline (2) was first isolated in 1991 by Bodo and co-workers from Malaysian medical plant Horsfildea superba Warb. (Myristicaceae) and used as a source of intoxicating snuffs.<sup>5</sup>

The unique 3,3-spirocyclic oxindole structural features with the quaternary center and potential biological

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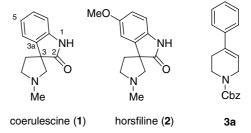


Figure 1.

oxidants promoted oxidative rearrangement (e.g., lead tetraacetate, 5b sodium tungstate, 7j tert-butylhypochlorite, <sup>7h</sup> and N-bromosuccinimide <sup>7b</sup>), Mannich reaction, <sup>7c</sup> ring-expansion reaction, 7f,i 1,3-dipolar [3+2] cycloaddition, 7d,g,l intramolecular radical cyclization, 7a,m-p and asymmetric nitroolefination reaction. 7e All of them find application on specific substrates for the construction of the 3,3-spirocyclic oxindole-derived alkaloids. Here, we report that a new synthetic study toward  $(\pm)$ coerulescine (1) and ( $\pm$ )-horsfiline (2) from 1-benzyloxycarbonyl-4-phenyl-1,2,5,6-tetrahydropyridine (3a) as shown in Figure 1.

# 2. Results and discussion

The retrosynthetic analysis of  $(\pm)$ -coerulescine (1) and  $(\pm)$ -horsfiline (2) is shown in Scheme 1. A remarkable

applications prompted the development of numerous strategies toward the synthesis of coerulescine (1) and horsfiline (2) from synthetic chemists.<sup>6,7</sup> Basically, the adopted strategies can be summarized in several

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coerulescine (1) 
$$\longrightarrow$$
  $\stackrel{N}{\longrightarrow}$   $\stackrel{N}{\longrightarrow}$   $\stackrel{CHO}{\longrightarrow}$  3a  $\stackrel{N}{\longrightarrow}$   $\stackrel{Cbz}{\longleftarrow}$   $\stackrel{Cbz}{\longleftarrow}$   $\stackrel{Cbz}{\longrightarrow}$  5a

Scheme 1.

two-step transformation is described as follows. One is the rapid access to produce 3-formyl-3-phenylpyrrolidine **5a** by rearrangement of 3,4-dihydroxy-4-phenylpiperidine. The other step is an intramolecular electrophilic cyclization<sup>8</sup> from 3-benzylcarbamoyl-3-phenylpyrrolidine to the 3,3-spirocyclic 2-oxindole skeleton **4** (a Fuji's intermediate of horsfiline).<sup>7e</sup>

The synthesis began from compounds 5a-c as shown in Scheme 2. Compounds 5a-c were prepared by six-step standard protocol from the commercially available 4hydroxypiperidine (6): (i) benzyloxycarbonylation of amine 6 with triethylamine and benzyloxycarbonyl chloride in tetrahydrofuran in ice bath for 1 h, (ii) oxidation of the resulting alcohol with Jones reagent in acetone in ice bath for 15 min, (iii) Grignard addition of ketone  $7^{9a}$ with different Grignard reagents (a,  $Ar = C_6H_5$ ; b,  $Ar = 4-FC_6H_4$ ; c,  $Ar = 3,4-CH_2O_2C_6H_3$ ) in tetrahydrofuran at -78 °C for 2 h, (iv) dehydration of the resulting tertiary alcohols with boron trifluoride etherate in dichloromethane in ice bath for 15 min, (v) dihydroxylation of olefins  $3\mathbf{a} - \mathbf{c}^{9b}$  with N-methylmorpholine N-oxide and a catalytic amount of osmium tetroxide in the cosolvent of tert-butanol and water at reflux temperature for 2 h,9c and (vi) rearrangement of diols 8a-c with boron trifluoride etherate in ice bath for 15 min. Thus, 3-aryl-3-formylpyrrolidines 5a-c could be obtained in good yield. The total synthetic procedure was monitored by TLC until the reaction was completed. The overall procedure was achieved within one working day.

While searching the related literature of 3-substituted 3-formylpyrrolidine, we found that it was a useful building

Scheme 2.

block in the synthesis of various potential biological compounds.  $^{10,11}$  In view of the experimental simplicity, we chose compound 3a as the model substrate to synthesize compound 5a in a moderate scale (0.1 mol). The simple two-step procedure of dihydroxylation and Lewis acid-catalyzed rearrangement reaction also provided enough amounts of material 5a in the synthesis of  $(\pm)$ -coerulescine (1) and  $(\pm)$ -horsfiline (2) in 73% yield.

When 3-benzyloxy-1-benzyloxycarbonyl-4-hydroxy-4-phenyl piperidine (8d) (prepared from monobenzylation of diol 8a) was treated with boron trifluoride etherate, the sole 3-formyl-3-phenylpyrrolidine 5a was still isolated in quantitative yield. The conversion of 8a–d to 5a–c is pinacol to pinacolone rearrangement. For providing aldehyde 5a, the similar reaction had been investigated on *N*-substituted 4-aryl-3,4-epoxypiperidines. According to the report, <sup>10a</sup> different *N*-substitutents (methyl or carbamate functional group) can affect the distribution of products.

With compound 5a in hand, aldehyde 5a was transformed into acid 9 by Jones oxidation as shown in Scheme 3. Treatment of acid 9 with dicyclohexylcarbodiimide and benzylamine gave a benzyl amide product. Intramolecular electrophilic cyclization<sup>8</sup> of the benzyl amide with tert-butylhypochlorite followed by zinc acetate gave the compound 4. The known approach had been investigated by Kikugawa and co-workers.8 Compound 4 was a known intermediate of horsfiline (2) in Fuji's report. <sup>7e</sup> According to this report, compound 4 could be transformed to horsfiline (2) via regioselective oxidation with lead tetrakis(trifluoroacetate) at the C-5 position and O-methylation of the resulting phenolic compound. This constitutes a formal synthesis of  $(\pm)$ horsfiline (2). Finally,  $(\pm)$ -coerulescine (1) was achieved by hydrogenation of compound 4 with hydrogen in the presence of a catalytic amount of 10% palladium on carbon followed by N-methylation with sodium borohydride and formaldehyde. The 1H NMR spectral data of compounds 1 and 4 were in accordance with those reported in the literature. <sup>7e,1</sup>

Scheme 3.

#### 3. Conclusion

We explored a Lewis acid catalyzed rearrangement reaction of 4-aryl-N-benzyloxycarbonyl-3,4-dihydroxypiperidine to form 3-aryl-3-formylpyrrolidine skeleton. This strategy of intramolecular electrophilic cyclization is synthetically useful for constructing ( $\pm$ )-coerulescine (1) and ( $\pm$ )-horsfiline (2) with 3,3-spirocyclic oxindole alkaloids. We are currently studying the scope of this process as well as additional applications of the methodology to the synthesis of piperidines, indolizidines, quinolizidines, and indoles.

# Supplementary data

Experimental procedures and photocopies of <sup>1</sup>H NMR (CDCl<sub>3</sub>) spectral data for 1, 3a–c, 4, 5a–c, 7, 8a–d, and 9 were supported. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2005.10.015.

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### References and notes

- (a) Cui, C. B.; Kakeya, H.; Osada, H. Tetrahedron 1996,
  52, 12651; (b) Cui, C. B.; Kakeya, H.; Osada, H. J. Antibiot. 1996, 49, 832.
- James, M. N. G.; Williams, G. J. B. Can. J. Chem. 1972, 50, 2407.
- 3. (a) Leclercq, J.; De Pauw-Gillet, M. C.; Bassleer, R.; Angenot, L. J. Ethnopharmacol. 1986, 15, 305; (b) Dupont, P. L.; Lamotte-Brasseur, J.; Dideberg, O.; Campsteyn, H.; Vermeire, M.; Angenot, L. Acta Crystallogr. Sect. B 1977, 33, 1801.
- (a) Finch, N.; Taylor, W. I. J. Am. Chem. Soc. 1962, 84, 1318; (b) Finch, N.; Taylor, W. I. J. Am. Chem. Soc. 1962, 84, 3871; (c) Shavel, J.; Zinnes, H. J. Am. Chem. Soc. 1962, 84, 1320.
- (a) Anderton, N.; Cockrum, P. A.; Colegate, S. M.; Edgar, J. A.; Flower, K.; Vit, I.; Willing, R. I. *Phytochemistry* 1998, 48, 437; (b) Colegate, S. M.; Anderton, N.; Edgar, J.; Bourke, C. A.; Oram, R. N. *Aust. Vet. J.* 1999, 77, 537; (c) Jossang, A.; Jossang, P.; Hadi, H. A.; Sévenet, T.; Bodo, B. *J. Org. Chem.* 1991, 56, 6527.
- For a review: Marti, C.; Carreira, E. M. Eur. J. Org. Chem. 2003, 2209.

- 7. (a) Jones, K.; Wilkinson, J. J. Chem. Soc., Chem. Commun. **1992**, 1767; (b) Pellegrini, C.; Strässler, C.; Weber, M.; Borschberg, H.-J. Tetrahedron: Asymmetry 1994, 5, 1979; (c) Bascop, S.-I.; Sapi, J.; Laronze, J.-Y.; Lévy, J. Heterocycles 1994, 38, 725; (d) Palmisano, G.; Annunziata, R.; Papeo, G.; Sisti, M. Tetrahedron: Asymmetry 1996, 7, 1; (e) Lakshmaiah, G.; Kawabata, T.; Shang, M.; Fuji, K. J. Org. Chem. 1999, 64, 1699; (f) Fischer, C.; Meyers, C.; Carreira, E. M. Helv. Chim. Acta 2000, 83, 1175; (g) Cravotto, G.; Giovenzana, G. B.; Pilati, T.; Sisti, M.; Palmisano, G. J. Org. Chem. 2001, 66, 8447; (h) Kuehne, M. E.; Roland, D. M.; Hafter, R. J. Org. Chem. 1978, 43, 3705; (i) Syam Kumar, U. K.; Ila, H.; Junjappa, H. Org. Lett. 2001, 3, 4193; (j) Somei, M.; Noguchi, K.; Yamagami, R.; Kawada, Y.; Yamada, K.; Yamada, F. Heterocycles 2000, 53, 7; (k) Bell, S. E. V.; Brown, R. F. C.; Eastwood, F. W.; Horvath, J. M. Aust. J. Chem. 2000, 53, 183; (1) Selvakumar, N.; Azhagan, A. M.; Srinivas, D.; Krishna, G. G. Tetrahedron Lett. 2002, 43, 9175; (m) Lizos, D. E.; Murphy, J. A. Org. Biomol. Chem. 2003, 1, 117; (n) Cossy, J.; Cases, M.; Pardo, D. G. Tetrahedron Lett. 1998, 39, 2331; (o) Lizos, D.; Tripoli, R.; Murphy, J. A. Chem. Commun. 2001, 2732; (p) Murphy, J. A.; Tripoli, R.; Khan, T. A.; Mali, U. M. Org. Lett. 2005, 7, 3287; (q) Kornet, T.; Thio, A. P. J. Med. Chem. 1976, 19, 892.
- (a) Prata, J. V.; Clemente, D. T. S.; Prabhakar, S.; Lobo, A. M.; Mourato, I.; Branco, P. S. J. Chem. Soc., Perkin Trans. 1 2002, 513; (b) Fleming, I.; Moses, R. C.; Tercel, M.; Ziv, J. J. Chem. Soc., Perkin Trans. 1 1991, 617; (c) Kikugawa, Y.; Kawase, M. J. Am. Chem. Soc. 1984, 106, 5728; (d) Kawase, M.; Kitamura, T.; Kikugawa, Y. J. Org. Chem. 1989, 54, 3394.
- (a) Hattori, K.; Sajiki, H.; Hirota, K. Tetrahedron 2000, 56, 8433;
  (b) Eastwood, P. R. Tetrahedron Lett. 2000, 41, 3705;
  (c) Bursavich, M. G.; West, C. W.; Rich, D. H. Org. Lett. 2001, 3, 2317;
  (d) Harriman, G. C. B.; Shao, J.; Luly, J. R. Tetrahedron Lett. 2000, 41, 8853.
- (a) Nagai, Y.; Hino, K.; Uno, H.; Minami, S. Chem. Pharm. Bull. 1980, 28, 1387; (b) Lyle, R. E.; Krueger, W. E. J. Org. Chem. 1965, 30, 394; (c) Hershenson, F. M.; Christensen, L. Synth. Commun. 1981, 11, 615.
- (a) Lyles-Eggleston, M.; Altundas, R.; Xia, J.; Sikazwe, D. M. N.; Pan, P.; Yang, Q.; Li, S.; Zhang, W.; Zhu, X.; Schmidt, A. W.; Vanase-Frawley, M.; Shrihkande, A.; Villalobos, A.; Borne, R. F.; Ablordeppey, S. Y. J. Med. Chem. 2004, 47, 497; (b) Xue, C.-B.; Chen, X.-T.; He, X.; Roderick, J.; Corbett, R. L.; Ghavimi, B.; Liu, R.-Q.; Covington, M. B.; Qian, M.; Ribadeneira, M. D.; Vaddi, K.; Trzaskos, J. M.; Newton, R. C.; Duan, J. J. W.; Decicco, C. P. Bioorg. Med. Chem. Lett. 2004, 14, 4453; (c) Hagen, S. E.; Domagala, J. M.; Heifetz, C. L.; Sanchez, J. P.; Solomon, M. J. Med. Chem 1990, 33, 849; (d) Jenkins, S. M.; Wadsworth, H. J.; Bromidge, S.; Orlek, B. S.; Wyman, P. A.; Riley, G. J.; Hawkins, J. J. Med. Chem. 1992, 35, 2392.